



# Contribution of protein conformational heterogeneity to NMR lineshapes at cryogenic temperatures

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While low-temperature Nuclear Magnetic Resonance (NMR) holds great promise for the analysis of unstable samples and for sensitizing NMR detection, spectral broadening in frozen protein samples is a common experimental challenge. One hypothesis explaining the additional linewidth is that a variety of conformations are in rapid equilibrium at room temperature and become frozen, creating an inhomogeneous distribution at cryogenic temperatures. Here, we investigate conformational heterogeneity by measuring the backbone torsion angle ( $\Psi$ ) in *Escherichia coli* Dihydrofolate Reductase (DHFR) at 105 K. Motivated by the particularly broad N chemical shift distribution in this and other examples, we modified an established NCCN  $\Psi$  experiment to correlate the chemical shift of  $N_{i+1}$  to  $\Psi_i$ . With selective  $^{15}\text{N}$  and  $^{13}\text{C}$  enrichment of Ile, only the unique I60-I61 pair was expected to be detected in  $^{13}\text{C}$ - $^{15}\text{N}$  correlation spectrum. For this unique amide, we detected three different conformation basins based on dispersed chemical shifts. Backbone torsion angles  $\Psi$  were determined for each basin:  $114 \pm 7^\circ$  for the major peak and  $150 \pm 8^\circ$  and  $164 \pm 16^\circ$  for the minor peaks as contrasted with  $118^\circ$  for the X-ray crystal structure (and  $118^\circ$  to  $130^\circ$  for various previously reported structures). These studies support the hypothesis that inhomogeneous distributions of protein backbone torsion angles contribute to the lineshape broadening in low-temperature NMR spectra.

solid state NMR | DNP | protein dynamics | backbone torsion angle | cryogenic protein studies

Many biophysical techniques, such as X-ray crystallography (1) and cryogenic electron microscopy (2), utilize low temperatures to reduce thermal motion and reduce irradiation damage during structure determination. The use of low temperatures has allowed investigations of systems that were previously unstable or inaccessible and in many cases improves the precision of structural studies. Decreasing the temperature of samples has also been shown to be beneficial in Nuclear Magnetic Resonance (NMR) experiments. At cryogenic temperatures, larger polarizations and improved detection sensitivities can be obtained. Low temperature also facilitates transfer of polarization from electron spins to nuclear spins (3) in a technique known as dynamic nuclear polarization (DNP) which can theoretically enhance the signal-to-noise ratio up to 660-fold.

In some cases, the use of low sample temperatures for NMR is accompanied by significant line broadening. Several hypotheses have been advanced to explain this lineshape. Solvent interactions were observed to correlate with the line broadening; specifically, the mean  $^{15}\text{N}$  linewidths in the DNP spectra of the Pfl bacteriophage were found to be different between hydrated (mean of 8 ppm and standard deviation (SD) of 1.4 ppm) vs. nonhydrated residues (mean of 6 ppm and SD of 1.7 ppm) (4). Alternatively, low-temperature broadening can result from exchange dynamics (5) or might result from the effects of the radicals used for DNP (6, 7), or from difficulties in optimizing cryogenic experimental protocols (8) including the use of relatively low spinning frequencies (4, 9). Several studies indicate that linewidths at low temperatures are typically primarily inhomogeneous, and many studies implicate conformational heterogeneity as an important source of line broadening in low-temperature NMR (10–16). In this hypothesis, rapid variation in various structural parameters occurs at room temperature due to conformational fluctuations, and measured chemical shifts and other properties are weighted averages of the ensemble. By contrast, freezing the sample results in a static inhomogeneous distribution of conformations and of chemical shifts. We explore this hypothesis to identify some of the conformational degrees of freedom that have broad distributions.

Of the many degrees of freedom in the protein conformation landscape, backbone torsion angles (17–19) are of particular interest and can be accessed by solid-state NMR (SSNMR) experiments. In this study, precise restraints for  $\Psi$  (19) are obtained through dephasing the  $\text{C}'$ - $\text{C}\alpha$  double-quantum coherence with  $^{15}\text{N}$  dipolar couplings. The experiment involves detecting the 2D double-quantum single-quantum (DQ-SQ)  $^{13}\text{C}$ - $^{13}\text{C}$  correlation spectrum and

## Significance

Understanding protein conformational flexibility is essential for insights into the molecular basis of protein function and the thermodynamics of proteins. Here, we investigate the ensemble of protein backbone conformations in a frozen solution of a protein, which may reflect (in restricted form) the ensemble in rapid equilibrium at room temperature. Various conformers of the backbone are spectrally resolved due to the exquisite sensitivity of Nuclear Magnetic Resonance (NMR) shifts to local conformations, and NMR methods allow us to directly probe the torsion angles corresponding to each band of chemical shifts.

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The authors declare no competing interest.

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recording the intensities of the cross-peaks as a function of C-N dephasing times. The sensitivity of this dephasing profile to the relative orientation of the C-N vectors allows a precise determination of torsion angles. This experiment, referred to as the “NCCN $\Psi$ ” measurement, has been initially demonstrated for crystalline ( $^{13}\text{C}$ ,  $^{15}\text{N}$ -Gly)-( $^{15}\text{N}$ -Gly)-Gly-HCl (17) and subsequently demonstrated on or applied to other peptides, such as MLF (20) and GNNQQNY fibrils from the yeast prion protein Sup35p (21), as well as SH3 protein residues (19) and  $\beta$ -hairpins in huntingtin exon1 fibrils (22). Here, we probe a backbone torsion angle  $\Psi$  at low temperature using a modified version of the “NCCN $\Psi$ ” measurement (17, 19) to correlate the dipolar angle restraint to the  $^{15}\text{N}$  shift.

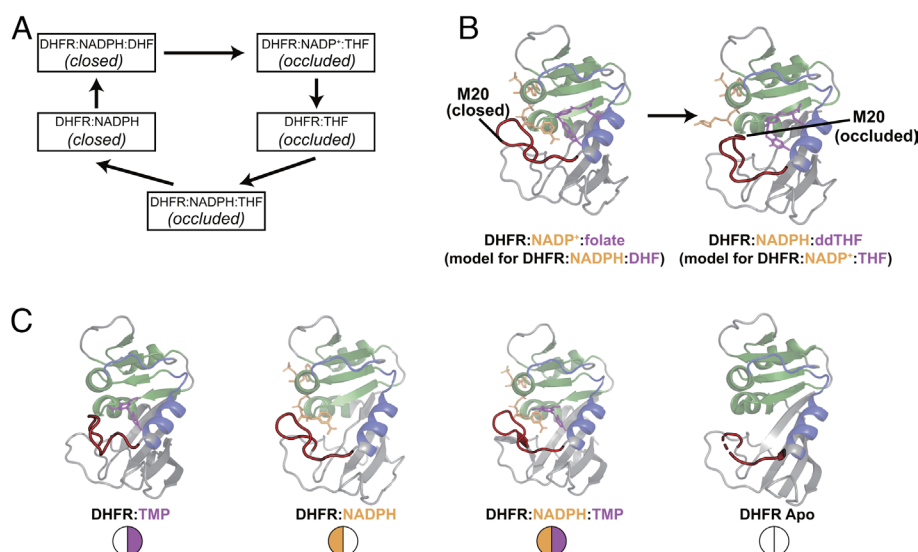
*Escherichia coli* DHFR is a globular enzyme that is very well studied from the point of view of conformational dynamics and is an important antibiotic target. DHFR catalyzes the reduction of dihydrofolate (DHF) to tetrahydrofolate (THF) using nicotinamide adenine dinucleotide phosphate (NADPH) as a cofactor. There are a number of intermediates in the catalytic cycle (Fig. 1A), including DHFR:NADPH, DHFR:NADPH:DHF, DHFR:NADP+:THF, DHFR:THF, and DHFR:NADPH:THF. To accommodate these chemical changes, DHFR accesses a variety of conformations, including two referred to as closed and occluded (Fig. 1B). Trimethoprim (TMP) is an effective antibiotic and has a very high binding affinity ( $K_d = 6$  pM) (23) to *E. coli* DHFR while competing for the substrate and appears to trap the protein in a closed conformation (Fig. 1C). The protein contains a cofactor binding pocket and a substrate binding pocket which both undergo intermediate exchange behavior at room temperature. A unique Ile-Ile amino acid pair (60-61) located in a beta-sheet structure is the subject of this study. The value of  $\Psi$  for residue 60 is  $118^\circ$  according to the X-ray crystallography structure of *E. coli* DHFR:TMP (24), and it varies from  $118^\circ$  to  $130^\circ$  among the structures of its native complexes in the catalytic cycle (25), as listed in [SI Appendix, Table S1](#).

## Results

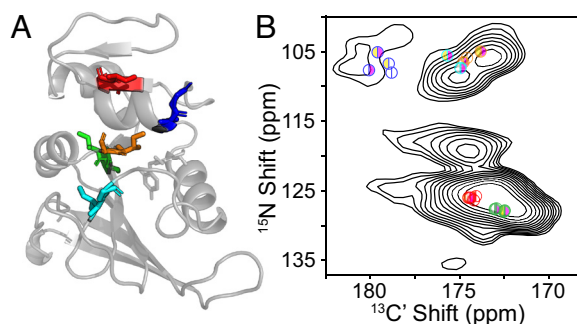
**Selective Isotopic Enrichment Scheme.** To explore the effects of conformational heterogeneity on lineshape, in view of the relatively

broad lines, we designed experiments involving selective isotopic enrichment that would highlight a mixture of putatively static and dynamic sites in  $^{13}\text{C}$ - $^{15}\text{N}$  correlation spectra. To that end, we prepared  $^{13}\text{C}$ ,  $^{15}\text{N}$ -Ile,  $^{15}\text{N}$ -Gly labeled *E. coli* DHFR:TMP (IG-DHFR), for which 5 sites will have  $^{13}\text{C}$ - $^{15}\text{N}$  directly bonded pairs detected in  $^{13}\text{C}$ ,  $^{15}\text{N}$  correlation spectrum, specifically a selective double cross-polarization (DCP) experiment with short contact time based on *E. coli* DHFR sequence: Ile2-Ser3 (sheet), Ile14-Gly15 (coil), Ile50-Gly51 (helix), Ile60-Ile61 (sheet), Ile94-Gly95 (coil), shown in Fig. 2A ( $^{15}\text{N}$ -Ser is present due to metabolic “scrambling” of  $^{15}\text{N}$ -Gly during protein expression). Several regions in *E. coli* DHFR are expected to be mobile in the mechanism and are in fact notably flexible based on comparing torsion angles in different DHFR ligand-bound structures. For example, sites in the M20 loop (especially Ile14 and Pro21) and the GH loop (including Ser148) vary significantly in their conformations for different complexes (24, 25) ([SI Appendix, Table S1](#)). Regarding residues observed in our experiment, the torsion angle variation of Ile14-Gly15 in the mobile M20 loop is up to  $187^\circ$ , and Ile14 is believed to flip during the transition from the closed to occluded conformations (I14 $\Psi = -11^\circ$  in DHFR:NADPH:DHF; I14 $\Psi = 157^\circ$  in DHFR:NADP+:THF) (25). In the DHFR:TMP complex, both Ile50 and Ile94 are close to the TMP ligand and are possibly affected by the flipping of the TMP tri-methoxybenzyl ring. By contrast, Ile2-Ser3 and Ile60-Ile61 are expected to be relatively immobile based on X-ray crystallography. Moreover, in support of this prediction, solution-state NMR  $T_2$  relaxation measurements of DHFR complexes and studies of its catalytic cycle, Gly15, Gly51, and Gly95 were observed to exhibit intermediate exchange behavior at room temperature (for the amidic NH group), while Ser3 and Ile61 were not (26). Using this isotope enrichment scheme, we explored whether there is any relation between the low-temperature linewidths and the previous indications for conformational dynamics.

In a separate experiment, we prepared  $^{13}\text{C}$ ,  $^{15}\text{N}$ -Ile labeled *E. coli* DHFR:TMP (I-DHFR), in which the only  $^{13}\text{C}$ - $^{15}\text{N}$  directly bonded pair expected to be detected in a selective  $^{15}\text{N}$ - $^{13}\text{C}$  correlation experiment is the amide connecting the only sequential I-I pair,



**Fig. 1.** (A) Conformational changes during the DHFR catalytic cycle (25). (A) DHFR catalyzes the reduction of DHF to THF. Five key complexes are proposed in the *E. coli* DHFR catalytic cycle. The DHFR:TMP state is considered analogous to DHFR:THF state (25). (B) The conformation of the M20 loop and other regions change from closed to occluded during hydride transfer. Residues that define the active site loop conformation, substrate-binding, and cofactor-binding markers are colored red, blue, and green, respectively. The substrates are colored magenta, and the cofactors are colored yellow. The design of the figures in (A) and (B) comes from previous work (25, 26). (C) The DHFR:TMP (T) (24) state and three associated DHFR liganded states are analyzed using solution-state NMR, including DHFR:NADPH (N) (PDB: 1rx1) state, DHFR:NADPH:TMP (NT) state, and the Apo (A) (PDB: 5dfr) state. Because the DHFR:NADPH:TMP (NT) state has no crystal structure available, a docking structure is shown with DHFR:NADPH (N) (PDB: 1rx1) as target and TMP as ligand structure.



**Fig. 2.** (A) Cartoon of the structure of *E. coli* DHFR:TMP (24) with pairs detected in low-temperature NMR spectra of IG-DHFR highlighted with stick rendering of the sidechains: I2-S3 (green), I14-G15 (cyan), I50-G51 (blue), I94-G95 (orange), and I60-I61 (red). (B) N-C' correlated DCP spectra of IG-DHFR with likely assignments indicated. The details of spectral collection and processing are listed in *SI Appendix, Table S2*. The chemical shift assignments for 4 liganded states of *E. coli* DHFR are shown with different markers as Fig. 1C and colored accordingly as (A). Significant overlap between these peaks introduces some ambiguity in these assignments as well as uncertainty in determining the lineshape of each individual site. Therefore, we next studied a sample where a single amide bond could be detected by low-temperature NMR.

residues 60–61. With this sample, we explored in detail the flexibility of a single site and the relation to low-temperature linewidths (without the complication of overlapping peaks due to other residues).

### Conformational Homogeneity in Solution-State NMR Spectra.

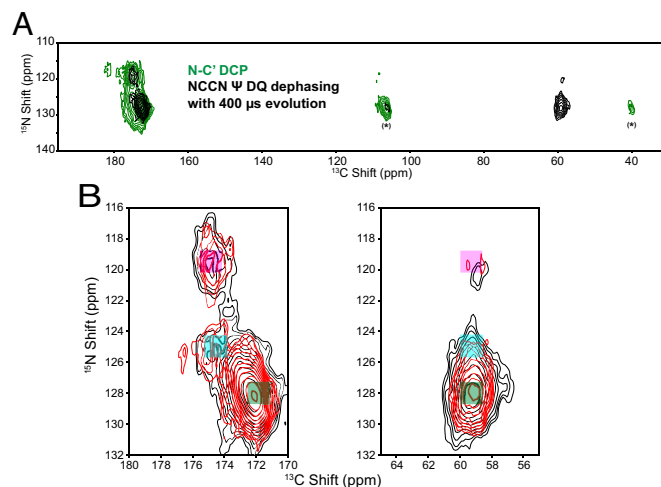
To prepare homogeneous *E. coli* DHFR samples in the desired ligand-bound state, we removed residual endogenous THF by an unfolding–refolding procedure after purification. Solution NMR spectra of refolded  $^{13}\text{C}$ ,  $^{15}\text{N}$ -Ile labeled *E. coli* DHFR:TMP (I-DHFR) (*SI Appendix, Fig. S2*) were compared to spectra from uniformly  $^{13}\text{C}$ ,  $^{15}\text{N}$  enriched samples from which assignments of 4 different liganded states were determined, including DHFR:TMP (T), DHFR:NADPH (N), DHFR:NADPH:TMP (NT), and Apo (A) state. All 12 Ile in the *E. coli* DHFR sequence were observed in the solution  $^{15}\text{N}$ -Heteronuclear Single Quantum Coherence (HSQC) spectra of I-DHFR. Chemical shifts and signal intensities match those expected based on the spectra of uniformly  $^{13}\text{C}$ ,  $^{15}\text{N}$  T state (*SI Appendix, Fig. S2A*), confirming successful selective enrichment and refolding. The same protein sample was used for low-temperature DNP-enhanced solid-state NMR (SSNMR) spectroscopy (Fig. 3 and *SI Appendix, Fig. S5*). Although the ligation of this I-DHFR sample is homogeneous (namely saturated with respect to TMP ligand and with no cofactor or other ligands bound), some Ile residues exhibit minor peaks suggesting a mixture of conformers. Ile61 (*SI Appendix, Fig. S2 B–D*), Ile155 (*SI Appendix, Fig. S2A*), and Ile41 (*SI Appendix, Fig. S2A*) have minor peaks with similar chemical shifts as either DHFR: NADPH: TMP (NT) state or DHFR: NADPH (N) state. Possibly, the minor peaks result from slow chemical exchange to other conformations.

**Lineshape Broadening in DNP-Enhanced NMR Spectra of Frozen Solutions of DHFR.** Five amide pairs are expected to appear in DNP-enhanced N-C correlation spectra of IG-DHFR, and 5 major peaks are indicated on the experimental data (Fig. 2B). Peak labels reflect putative assignments based on comparison to solution-state NMR chemical shifts, although peak overlap and minor additional peaks cause these to be tentative assignments. The  $^{15}\text{N}$  amide linewidth (rmsd) of these residues varies from 3.4 ppm to 8.2 ppm (*SI Appendix, Fig. S3*), while the  $^{13}\text{C}'$  linewidth varies from 1.9 ppm to 2.3 ppm. There is no apparent relationship between the magnitude of linewidth at low temperature and indications of chemical shift variation due to conformational

dynamics at room temperature from solution-state NMR studies (*SI Appendix, Fig. S4A*). Several  $^{15}\text{N}$  linewidths are considerably larger than the range in chemical shift expected for the various species in exchange in solution (less than 2 ppm) (26), and the two quantities are apparently uncorrelated. Thus, the variation in isotropic shift observed in DNP is probably not specifically due to the millisecond and microsecond conformational exchange processes observed in solution NMR in this example.

**Torsion Angle Measurements.** In order to explore the conformational heterogeneity within a single confidently assigned amino acid residue, we characterized refolded  $^{13}\text{C}$ ,  $^{15}\text{N}$ -Ile labeled *E. coli* DHFR:TMP (I-DHFR) using chemical shift and torsion angle measurements, probing the unique I-I pair in DHFR (60–61). To test whether the broad lines in the low-temperature spectra of DHFR reflect an inhomogeneous mixture of various conformers, restraints on the torsion angle  $\Psi$  were obtained using the NCCN  $\Psi$  double quantum (DQ) dephasing experiment (17, 19) (illustrated in *SI Appendix, Fig. S1*), modified to be a pseudo-3D experiment, with chemical shifts for N and C' measured in a 2D spectrum, and dephasing for torsion angle determination in the third dimension.

We investigated whether distinct chemical shifts in the broad low-temperature peak are associated with distinct conformations by comparing the dephasing profiles at various positions in the N-C correlation spectrum. The 2D N-C' correlation spectrum is shown in Fig. 3A (green). The chemical shift range of I61N is 115 ppm to 135 ppm, and the I60C' chemical shift range is 170 ppm to 180 ppm. The extremely broad peak of I60C'-I61N appears to be organized in multiple “basins,” suggesting the existence of various conformations (collections of similar structures). The N-C correlation spectrum with the shortest evolution time is shown in Fig. 3 (black). The control experiment,  $S_0$  without dephasing (Fig. 3A, black) is compared to the simple double cross-polarization (DCP) 2D spectrum (Fig. 3A, green) and has a similar peak shape with approximately 15% of the signal intensity since losses occur



**Fig. 3.** DNP spectra of  $^{13}\text{C}$ ,  $^{15}\text{N}$ -Ile DHFR:TMP (I-DHFR) collected at 105 K. (A) The N-C' correlation spectrum of I-DHFR (green) collected with double cross-polarization (DCP) pulse sequence (27, 28) is compared with the  $S_0$  spectrum with 400  $\mu\text{s}$  evolution ( $t_{\text{CN}} = 4 \pi$ ) of the NCCN  $\Psi$  double quantum (DQ) dephasing experiment (black). No  $C\alpha$  intensity is observed after the DCP transfer (green), which is spectrally selective for  $N_{\alpha+1}$  to  $C_i$ . The asterisks (\*) indicate the first and second spinning sidebands for the  $C'$  resonance. In the NCCN  $\Psi$  DQ dephasing experiment, the  $^{13}\text{C}\alpha_i$ - $^{13}\text{C}'_i$  DQ coherence was reconverted to both  $C\alpha$  and  $C'$  with SPC5 (29), resulting in the observed  $C\alpha$  peaks (black). (B) The zoomed-in  $S_0$  spectra (black) and  $S_\alpha$  spectra (red) with 400  $\mu\text{s}$  evolution at  $NC'$  and  $NC\alpha$  regions. Three peaks at N chemical shifts of 128 ppm, 125 ppm, and 119.5 ppm are highlighted with green (peak 1), cyan (peak 2), and purple (peak 3) square overlaid ornaments. The details of spectral collection and processing are listed in *SI Appendix, Table S2*.



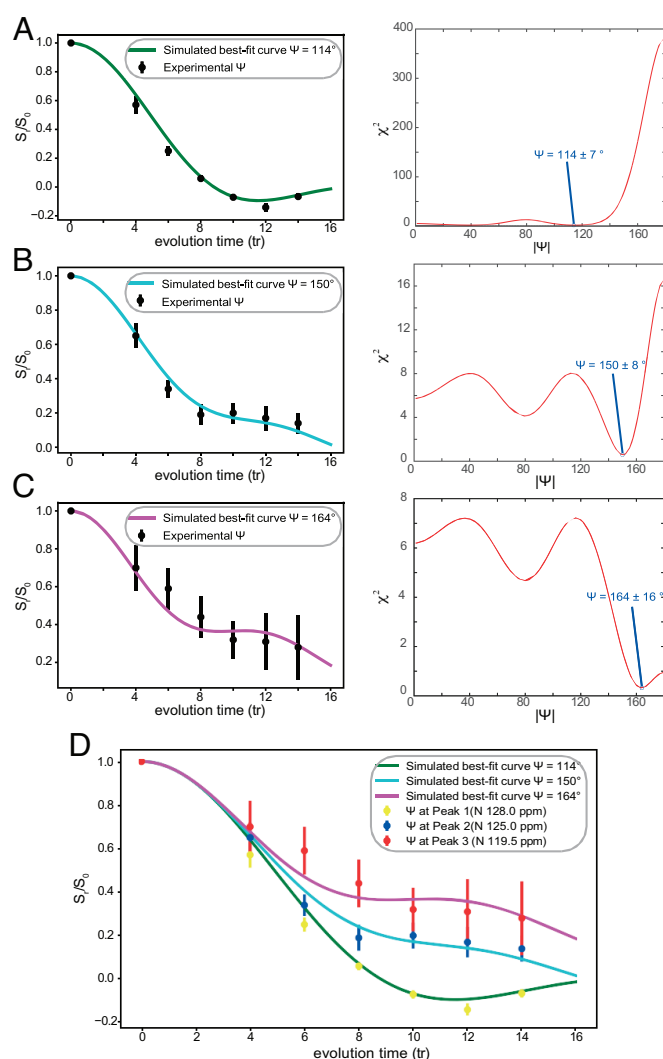
when exciting double quantum coherences and then reconverting back to single quantum coherence. Considering the peak shape of I60C-I61N, three peak regions were defined for torsion angle analysis (Fig. 3B): peak 1 ( $^{15}\text{N}$  128 ppm,  $^{13}\text{C}'$  172.5 ppm,  $^{13}\text{C}\alpha$  59.5 ppm, green), peak 2 ( $^{15}\text{N}$  125 ppm,  $^{13}\text{C}'$  175 ppm,  $^{13}\text{C}\alpha$  59.5 ppm, cyan), and peak 3 ( $^{15}\text{N}$  119.5 ppm,  $^{13}\text{C}'$  175.5 ppm,  $^{13}\text{C}\alpha$  59.5 ppm, purple). The normalized intensity ratios of each dephased spectrum (pulse sequence *SI Appendix*, Fig. S1C) and its corresponding reference spectrum (pulse sequence *SI Appendix*, Fig. S1D) ( $S_r/S_0$  defined in equation 2 in *SI Appendix*) were extracted from all the 2D spectra with evolution times varying from  $4\tau_r$  to  $14\tau_r$  (where  $\tau_r$  represents the duration of a rotor period, calculated as  $1/n_r$ ) (*SI Appendix*, Fig. S5). The intensities are plotted as a dephasing curve for torsion angle analysis (Fig. 4), with each of the three “subpeaks” of I60 of DHFR:TMP indicated separately (dephasing times as listed in the figure caption). The data from the NCCN  $\Psi$  DQ dephasing experiment indicate that at least three subpeaks in the I60 spectrum have different dephasing time profiles. Using a Chi-square test to fit the three peaks, there appear to be three conformations present with  $\Psi = 114 \pm 7^\circ$ ,  $150 \pm 8^\circ$ , and  $164 \pm 16^\circ$ , respectively. The error bar of the Chi-square fit is determined by a 90% confidence level. Details of the NCCN  $\Psi$  DQ dephasing pulse sequences and curve fittings are included in *SI Appendix*.

X-ray crystallography of *E. coli* DHFR: TMP shows that Ile60 and Ile61 are located in a beta-sheet with  $\Psi$  for I60 of  $118^\circ$ , which is within the error bar range of peak 1. The X-ray structures of *E. coli* DHFR complexed in various liganded states show that the value of I60 $\Psi$  varies from  $118^\circ$  to  $130^\circ$ . The two complexes with the closed conformation in *E. coli* DHFR catalytic cycle exhibit a  $\Psi$  angle of  $120^\circ$  and  $122^\circ$  at Ile60.

Despite the fact that the range of torsion angle values indicated by these SSNMR data exceeds that in the crystal structures, it is noteworthy that the various angles are all compatible with the idea that the residue remains as a beta sheet (remains in the “beta basin”). The range of chemical shifts also suggests beta structures [based on PLUQ (30)]. The three peaks observed at low temperature have  $\text{C}'$  and  $\text{C}\alpha$  chemical shifts compatible with typical sheet or random coil values (31). The average N chemical shift for Ile in beta-sheet conformations is  $124 \pm 5$  ppm, while that for a random coil is  $121 \pm 5$  ppm (31), indicating that the two stronger peaks (peak 1 and peak 2 with N chemical shift of 125 ppm and 128 ppm) are likely to be beta-sheet structures. Peak 1 has a N shift of 119.5 ppm, which is compatible with random coil or sheet.

The average properties indicated by crystallography or solution NMR agree well with the average of the three peaks seen at low temperature. Weighted averages of the torsion angle values determined for the three low-temperature SSNMR peaks are in the range of the values from crystallography ( $118^\circ$ ) and a typical value for beta sheets. The average of the chemical shifts in the low-temperature SSNMR data, using weighted sum of the three peaks, were also in good agreement with the solution NMR shifts. *SI Appendix*, Table S4 analyses three shifts in this fashion: Ile61N, Ile60C', and Ile60C $\alpha$ . Overall, it is reasonable to hypothesize that the solution ensemble includes a rapid average of three states corresponding to the three peaks observed at low temperature.

We eliminated the possibility of irreversible sample alteration during DNP sample preparation and freezing. After the low-temperature experiments, the DNP sample was recovered, buffer exchanged, and characterized with room-temperature solution-state NMR experiments. Comparing the  $^{15}\text{N}$ -HSQC spectra before and after DNP measurements (*SI Appendix*, Fig. S6), all 12 Ile residues were observed and preserved at the same peak positions and approximate relative intensities. The sample concentration calculation based on



**Fig. 4.** Dipolar dephasing of (A) peak 1 (N 128.0 ppm, green in Fig. 3B), (B) peak 2 (N 125.0 ppm, cyan in Fig. 3B), and (C) peak 3 (N 119.5 ppm, magenta in Fig. 3B) with corresponding best-fit curves based on a Chi-square test. The normalized intensity ratio  $S_r/S_0$  was calculated from peak intensities in  $S_0$  and  $S_r$  spectra (*SI Appendix*, Fig. S5) collected using the NCCN  $\Psi$  DQ dephasing experiment (17, 19) (illustrated in *SI Appendix*, Fig. S1). The overlap of the three decay curves is displayed in (D), which clearly shows the different decay time profiles for the three peaks.

the spectral intensity indicates that 85% of the protein sample was successfully recovered for this analysis, and we conclude that the structures we observed at the low temperature result from reproducible conformational basins reversibly trapped by freezing, rather than degradation of the protein.

Since peak 1 (with  $^{15}\text{N}$  chemical shift of 128 ppm) is intense and broad, we asked whether there exist multiple conformations within this broad peak, each with its own torsion angle. The analysis is shown in *SI Appendix*, Fig. S7. Three peak regions were picked based on the peak shape of the  $S_r$  spectra after 400  $\mu\text{s}$  (*SI Appendix*, Fig. S7): peak 1A (N 127 ppm), peak 1B (N 129 ppm), and peak 1C (N 130 ppm). Based on Chi-square fits, peak 1A is associated with  $\Psi = 115 \pm 8^\circ$  while peak 1B and peak 1C are associated with  $\Psi = 116^\circ$  and  $115^\circ$ , but in both cases, the minimum chi squared value exceeded the range for the 90% CI (Chi-square of peak 1B is 5.3 and peak 1C is 3.3), indicating that neither peak 1B nor peak 1C is compatible with a different torsion angle value. This result indicates that this broad peak is likely to be associated with a constant torsion angle  $\Psi$ , and the line

broadening within peak 1 may be associated with other structural or spectroscopic parameters.

## Discussion

The torsion angle variation observed for residue 60 at cryogenic temperatures has a larger range than that reported in X-ray structures including different ligated states. Such a variation was not anticipated based on the thermal disorder in the structures, since the X-ray crystallography structures of either closed (PDB: 1rx2) or occluded (PDB: 1rx6) conformations of *E. coli* DHFR report B-factors smaller than 15 Å<sup>2</sup> for backbone atoms in I60-I61 (25) suggesting good precision and rigidity. From the point of view of millisecond and microsecond dynamics and line broadening in solution NMR at room temperature, I60 is not expected to be an exceptionally flexible residue (26, 32), without pronounced relaxation dispersion observed in native ligated states. Thus, by all indications, it is a structured residue within a beta-strand (24–26). Notably, low-temperature linewidths for this residue are comparable to those from many other residues (7) and other samples (4, 33, 34). Therefore, we hypothesize that structural variations such as large backbone torsion angles likely occur for numerous other backbone sites, while the protein remains stably folded. One possible interpretation is that the variation in (or interconversion between) peaks 1, 2, and 3 also involves correlated motions of other degrees of freedom within the protein, for example,  $\Phi$  of the succeeding residue, so that overall the backbone shape is not significantly displaced. In this case, a “rocking” motion of the amide plane within a stably folded protein would occur. Similar motions of the backbone have been discussed previously based on NMR and dynamics simulations (35, 36) and in terms of their significance for protein structural determination (29, 37).

The timescale of interconversion between structures associated with peak 1, peak 2, or peak 3 is not known directly from these data. The narrow solution NMR linewidths for this residue (and others) observed at room temperature suggest that these structures (corresponding to peaks 1, 2, and 3) interconvert on a timescale that cannot be captured by CPMG experiments. Fast timescale

backbone and sidechain dynamics of DHFR (~50 ps to 10 ns) have been studied using solution NMR indicating  $S^2 > 0.8$  for the NH vector (38). This relatively rigid description was determined to be consistent with the diffraction data for the crystalline protein (39). The present cryogenic SSNMR data might suggest a comparable or somewhat lower-order parameter for Ile60. The data might suggest a two-site model with 30% of the population at an angle of 40°, giving  $S^2 \sim 0.65$ ; however, integrated intensities for these broad peaks are inaccurate, and the freezing protocol could be expected to result in an ensemble that is different to room temperature. Possibly, SSNMR data may reflect dynamics on different timescales (sub ps) than solution NMR consistent with some previous studies (35, 36). Inhomogeneously broad lines in cryogenic NMR spectra suggest that the structures interconvert on a timescale of many ms or slower at 105 K. If these conjectures are correct, a very steep change in the rate constant with respect to the temperature suggests that the solvent (and its phase change) plays an important role.

In conclusion, we determined the range of backbone torsion angle  $\Psi$  for a specific residue in a frozen protein, DHFR, at 105 K using a modified NCCN solid-state NMR dihedral angle measurement. We showed that the broad NMR peak for the single amide N-C' correlation of I60-I61 contains multiple conformational subspecies with distinct chemical shifts and torsion angles. This result supports the hypothesis that the origin of broad lines in NMR of frozen samples is dominated by conformational heterogeneity, including backbone torsion angle heterogeneity, at least for some protein sites.

**Data, Materials, and Software Availability.** NMR data have been deposited in Biological Magnetic Resonance Data Bank (BMRB Entry 52235) (40, 41).

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